

Diastereo- and Enantioselective Construction of 3,3'-Pyrrolidinyldispirooxindole Framework via Catalytic Asymmetric 1,3-Dipolar Cycloadditions

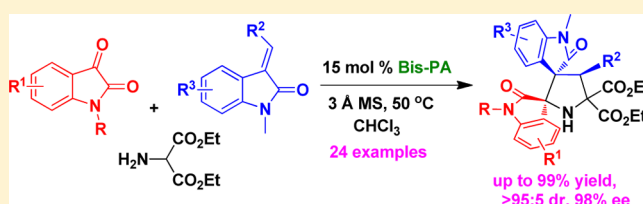
Wei Dai,[†] Xiao-Li Jiang,[†] Qiong Wu,[‡] Feng Shi,^{*,†} and Shu-Jiang Tu[†]

[†]Jiangsu Key Laboratory of Green Synthetic Chemistry for Functional Materials, School of Chemistry & Chemical Engineering, Jiangsu Normal University, Xuzhou 221116, China

[‡]School of Chemistry and Chemical Engineering, Xuzhou Institute of Technology, Xuzhou 221008, China

S Supporting Information

ABSTRACT: The first catalytic enantioselective construction of a 3,3'-pyrrolidinyldispirooxindole scaffold has been established via organocatalytic asymmetric 1,3-dipolar cycloadditions of isatin-derived azomethine ylides with methyl-eneindolinones, which afforded structurally complex bis-spirooxindoles containing three contiguous and two quaternary stereogenic centers in generally high yields (up to 99%) and excellent diastereo- and enantioselectivities (up to >95:5 dr, 98% ee). This reaction also provides a good example for the application of catalytic asymmetric 1,3-dipolar cycloadditions in constructing enantioenriched bis-spirooxindole frameworks with structural complexity and rigidity.



INTRODUCTION

Spirooxindoles are fascinating frameworks which constitute the core structures of many natural alkaloids and bioactive molecules.^{1,2} Among them, 3,3'-pyrrolidinyldispirooxindole skeletons are frequently found in natural products as exemplified by (–)-horsifiline^{1c} and (–)-spirotryprostatin A^{1d} (Figure 1). More

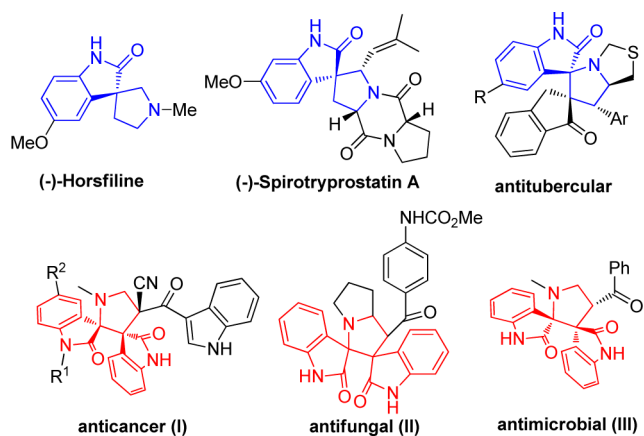


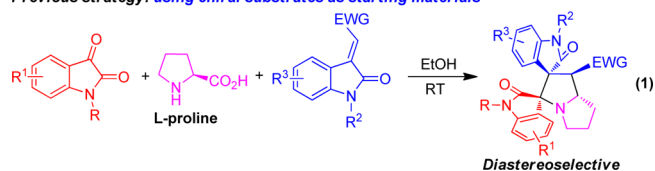
Figure 1. Some 3,3'-pyrrolidinyldispirooxindoles and bis-spirooxindoles.

importantly, 3,3'-pyrrolidinyldispirooxindoles (compounds I–III) exhibit prominent bioactivities such as anticancer,^{2a} antifungal,^{2b} and antimicrobial^{2c} activities. As a result, it is highly valuable to construct this class of bis-spirooxindole scaffolds, especially in an enantioselective manner, because the

bioactivity of the two enantiomers may differ from each other or from the racemic compounds.³

However, a survey of the literature disclosed that most of the approaches were concentrated on the synthesis of racemic bis-spiro products,⁴ and no enantioselective methods have been reported to access this type of chiral bis-spirooxindoles.⁵ This is mainly ascribed to the difficulty in constructing such structurally rigid bis-spirooxindole frameworks with at least two quaternary stereogenic centers. Recently, Chen, Yang, and co-workers^{5a} and the Shi group^{5b} developed a diastereoselective strategy to achieve the construction of a chiral 3,3'-pyrrolidinyldispirooxindole skeleton by using chiral L-proline as a starting material (eq 1). In spite of these creative works,⁵

Previous strategy: using chiral substrates as starting materials



the catalytic enantioselective construction of this class of bis-spirooxindole scaffolds by utilizing achiral starting materials has not yet been achieved. Thus, there is a great demand to develop catalytic asymmetric strategies for the construction of such bis-spirooxindole frameworks in an enantioselective way.

One of the most efficient ways to enantioselectively construct pyrrolidine skeleton is Lewis acid catalyzed⁶ or organocatalytic⁷

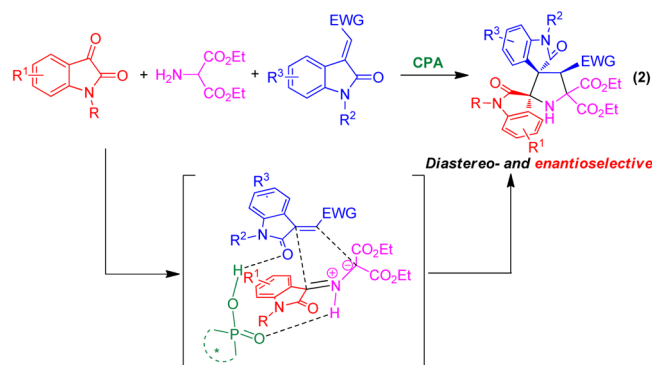
Received: March 31, 2015

Published: May 4, 2015

asymmetric 1,3-dipolar cycloaddition (1,3-DC) of azomethine ylide.⁸ In particular, chiral phosphoric acid⁹ (CPA) catalyzed asymmetric 1,3-DC of isatin-derived azomethine ylide with electron-deficient alkenes or alkynes has proven to be a robust method to build up the 3,3'-pyrrolidinylspirooxindole scaffold in a highly enantioselective fashion.¹⁰ However, this strategy has not been employed for the enantioselective construction of the structurally more complex and rigid bis-spirooxindole motif, which remains a great challenge in catalytic asymmetric 1,3-DCs. In order to confront this challenge, we designed a CPA-catalyzed asymmetric 1,3-DC of isatin-derived azomethine ylide with methyleneindolinone (Scheme 1, eq 2). On the basis of

Scheme 1. Design of Catalytic Asymmetric 1,3-DC to Enantioselectively Construct the 3,3'-Pyrrolidinylspirooxindole Framework

This strategy: catalytic asymmetric approaches using achiral starting materials



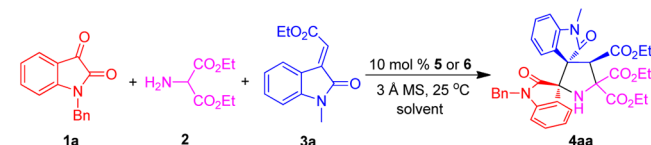
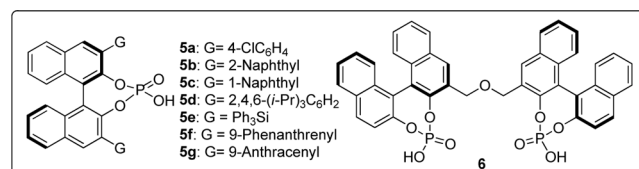
our previous experience in CPA-catalyzed reactions,¹¹ we envisioned that the two reactants would be simultaneously activated by CPA via a dual hydrogen-bonding activation mode, thus leading to the diastereo- and enantioselective construction of the 3,3'-pyrrolidinylspirooxindole framework (Scheme 1).

Herein, we report the first diastereo- and enantioselective construction of the 3,3'-pyrrolidinylspirooxindole scaffold via a catalytic asymmetric 1,3-DC of isatin-derived azomethine ylide with methyleneindolinone, which efficiently afforded structurally complex bis-spirooxindoles bearing two quaternary and three contiguous stereogenic centers in high yields (up to 99% yield) and excellent diastereo- and enantioselectivities (up to >95:5 dr, 98% ee).

RESULTS AND DISCUSSION

At the beginning, the reaction of *N*-benzylisatin **1a**, diethyl 2-aminomalonate **2**, and methyleneindolinone **3a** was employed as a model reaction to evaluate BINOL-derived CPAs **5a–g** (Table 1, entries 1–7). Although these reactions afforded the desired bis-spirooxindole product **4aa** in high yields (79%–86%) and excellent diastereoselectivities (all >95:5 dr), the enantioselective control of these catalysts on the reaction was very frustrating. In detail, CPAs **5a–f** bearing either small or bulky 3,3'-substituents nearly had no enantioselective induction on the reaction, and the generated bis-spirooxindole products were almost racemates (entries 1–6). Only CPA (**5g**) with a very bulky 9-anthracenyl group at the 3,3'-positions of the BINOL-backbone could control the enantioselectivity to some extent, but the ee value was still very low (entry 7). These results indicated that the capability of BINOL-derived CPAs in controlling the enantioselectivity of this reaction was not promising. In recent years, chiral bis-phosphoric acid (Bis-PA,

Table 1. Screening of Catalysts and Solvents^a

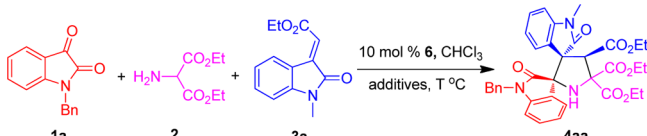


entry	solvent	Cat.	yield ^b (%)	dr ^c	ee ^d (%)
1	toluene	5a	83	>95:5	0
2	toluene	5b	85	>95:5	4
3	toluene	5c	80	>95:5	5
4	toluene	5d	86	>95:5	6
5	toluene	5e	79	>95:5	3
6	toluene	5f	83	>95:5	3
7	toluene	5g	81	>95:5	31
8	toluene	6	61	>95:5	68
9	CH ₂ Cl ₂	6	39	>95:5	57
10	CHCl ₃	6	56	>95:5	82

^aUnless indicated otherwise, the reaction was carried out on a 0.1 mmol scale in a solvent (1 mL) with 3 Å MS (100 mg) at 25 °C for 15 h, and the mole ratio of **1a**/2/**3a** was 1.2:1:1.5. ^bIsolated yield. ^cThe diastereomeric ratio (dr) was determined by ¹H NMR and HPLC. ^dThe enantiomeric excess (ee) was determined by HPLC.

6) has exhibited its great potential in catalytic asymmetric 1,3-DCs,^{7c,10b,d} which is far superior to mono-CPAs in terms of enantioselective control. Therefore, we tentatively utilized Bis-PA (**6**) as a catalyst to the model reaction. Indeed, this reaction under the catalysis of Bis-PA (**6**) delivered the product **4aa** in a much higher enantioselectivity of 68% ee albeit with a moderate yield of 61% (entry 8). Then, in the presence of Bis-PA (**6**) as the most suitable catalyst, the model reaction was subjected to a variety of solvents mainly including arenes and chloro-substituted alkanes (see the Supporting Information for details) with the aim of further improving the enantioselectivity. It was found that chloroform could offer the reaction in the highest enantioselectivity of 82% ee, although the yield was decreased to some extent (entry 10). Thus, chloroform was selected as an optimal reaction medium for subsequent condition optimization.

As shown in Table 2, further optimization of reaction conditions was focused on screening additives, temperature, reaction time, and catalyst loading. The evaluation of molecular sieves (MS) and anhydrous sulfates as water absorbers disclosed that anhydrous sulfates were inferior to MS with regard to the enantioselectivity (entries 4–5 vs 1–3). Among different types of MS, 4 Å MS afforded the best enantioselectivity of 87% ee, but the yield was very low (entry 2). Considering the balance of the yield and the enantioselectivity, 3 Å MS was finally chosen as a more suitable reaction additive (entry 1). Then, by elongating the reaction time from 15 to 36 h, the yield was improved from 56% to 74% with almost retained enantioselectivity (entry 1 vs 6). Furthermore, elevating the reaction temperature from 25 to 50 °C resulted in a significantly increased yield of 94% and a slightly ameliorated enantioselectivity of 83% ee (entry 7 vs 6). Finally, properly increasing the catalyst loading to 15 mol % led

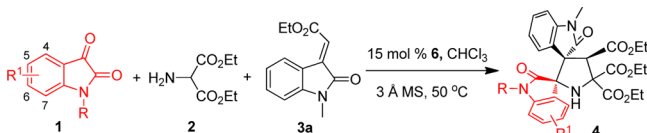
Table 2. Further Optimization of Conditions^a


entry	additives	T (°C)	yield ^b (%)	dr ^c	ee ^d (%)
1	3 Å MS	25	56	>95:5	82
2	4 Å MS	25	33	>95:5	87
3	5 Å MS	25	44	>95:5	82
4	MgSO ₄	25	61	>95:5	55
5	Na ₂ SO ₄	25	59	>95:5	57
6 ^e	3 Å MS	25	74	>95:5	80
7 ^e	3 Å MS	50	94	>95:5	83
8 ^{e,f}	3 Å MS	50	99	>95:5	86

^aUnless indicated otherwise, the reaction was carried out on a 0.1 mmol scale in chloroform (1 mL) with additives (100 mg) at the indicated temperature for 15 h, and the mole ratio was 1a/2/3a was 1.2:1:1.5. ^bIsolated yield. ^cThe diastereomeric ratio (dr) was determined by ¹H NMR and HPLC. ^dThe enantiomeric excess (ee) was determined by HPLC. ^eThe reaction time was 36 h. ^fIn the presence of 15 mol % of 6.

to a quantitative yield of 99% and a high enantioselectivity of 86% ee (entry 8).

After establishing the optimal reaction conditions, we carried out the investigation on the substrate scope of isatins **1** via the reactions with diethyl 2-aminomalonate (**2**) and methyleneindolinone **3a**. As shown in Table 3, a wide scope of isatins bearing different R/R¹ groups could be successfully utilized to the catalytic asymmetric 1,3-DCs, which offered the desired bis-spirooxindoles **4** with two quaternary stereogenic centers in high yields (75–99%) and excellent stereoselectivities (all >95:5 dr, 82–92% ee). In detail, this protocol is applicable to a series of *N*-benzyl-, phenyl-, or alkyl-substituted isatins (entries

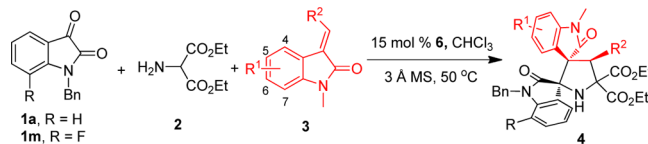
Table 3. Substrate Scope of Isatins 1^a


entry	4	R/R ¹ (1)	yield ^b (%)	dr ^c	ee ^d (%)
1	4aa	Bn/H (1a)	99	>95:5	86
2	4ba	<i>p</i> -tBuC ₆ H ₄ CH ₂ /H (1b)	95	>95:5	88
3	4ca	<i>p</i> -BrC ₆ H ₄ CH ₂ /H (1c)	89	>95:5	84
4	4da	<i>m</i> -ClC ₆ H ₄ CH ₂ /H (1d)	85	>95:5	88
5	4ea	<i>o</i> -ClC ₆ H ₄ CH ₂ /H (1e)	86	>95:5	86
6	4fa	Ph/H (1f)	88	>95:5	90
7	4ga	Me/H (1g)	83	>95:5	80
8	4ha	Bn/5-F (1h)	93	>95:5	86
9	4ia	Bn/6-MeO (1i)	75	>95:5	82
10	4ja	Bn/6-Cl (1j)	90	>95:5	90
11	4ka	Bn/6-Br (1k)	90	>95:5	88
12	4la	Bn/7-CF ₃ (1l)	82	>95:5	88
13	4ma	Bn/7-F (1m)	87	>95:5	92

^aUnless indicated otherwise, the reaction was carried out on a 0.1 mmol scale in chloroform (1 mL) with 3 Å MS (100 mg) at 50 °C for 36 h, and the mole ratio of 1:2:3a was 1.2:1:1.5. ^bIsolated yield. ^cThe diastereomeric ratio (dr) was determined by ¹H NMR. ^dThe enantiomeric excess (ee) was determined by HPLC.

1–7), and there is no significant difference among *N*-benzyl-substituted substrates **1a–e** in terms of enantioselectivity (entries 1–5). Notably, *N*-phenylisatin **1f** displayed higher capability in enantioselective control than its *N*-benzyl- or *N*-alkyl-substituted counterparts (entry 6 vs 1–5 and 7). In addition, a variety of isatins with different substituents at the C5–C7 positions of the phenyl ring could smoothly participate in the reaction in generally high yields and good enantioselectivities (entries 8–13). It seemed that the position of the substituent had some effect on the enantioselectivity since C7-fluoro-substituted isatin delivered a higher ee value than the C5-fluoro-substituted isatin (entry 13 vs 8). The electronic nature of the substituents might also influence the enantioselectivity to some extent because substrates **1j,k** offered the products with higher ee value than substrate **1i** did (entries 10 and 11 vs 9).

Next, the substrate scope with respect to methyleneindolinones **3** was studied by the reactions with isatin **1a** or **1m** and amino ester **2** under the standard conditions. As illustrated in Table 4, this approach was amenable to a wide range of

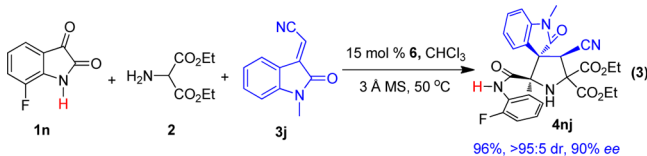
Table 4. Substrate Scope of Methyleneindolinones 3^a


entry	4	R (1)	R ¹ /R ² (3)	yield ^b (%)	dr ^c	ee ^d (%)
1	4ab	H (1a)	5-Me/CO ₂ Et (3b)	90	>95:5	88
2	4mc	F (1m)	5-Cl/CO ₂ Et (3c)	64	>95:5	86
3	4md	F (1m)	6-MeO/CO ₂ Et (3d)	90	>95:5	92
4	4mae	F (1m)	6-Br/CO ₂ Et (3e)	87	>95:5	94
5	4mf	F (1m)	6-Cl/CO ₂ Et (3f)	85	>95:5	92
6	4ag	H (1a)	7-Me/CO ₂ Et (3g)	82	>95:5	88
7	4mh	F (1m)	7-F/CO ₂ Et (3h)	62	>95:5	86
8	4mi	F (1m)	H/COPh (3i)	80	>95:5	98
9	4mj	F (1m)	H/CN (3j)	92	>95:5	96

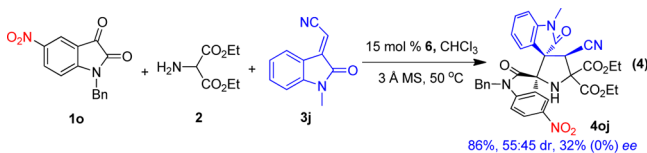
^aUnless indicated otherwise, the reaction was carried out on a 0.1 mmol scale in chloroform (1 mL) with 3 Å MS (100 mg) at 50 °C for 36 h, and the mole ratio of 1/2/3 was 1.2:1:1.5. ^bIsolated yield. ^cThe diastereomeric ratio (dr) was determined by ¹H NMR. ^dThe enantiomeric excess (ee) was determined by HPLC.

methyleneindolinones bearing distinct R¹/R² groups, leading to structurally complex bis-spirooxindoles in overall high yields (62%–92%) and excellent diastereo- and enantioselectivities (all >95:5 dr, 86%–98% ee). For the R¹ group, there was no obvious electronic effect on the enantioselectivity because either electron-donating or electron-withdrawing groups at the same position could give the similar ee value (entry 1 vs 2, entry 3 vs 4–5, entry 6 vs 7), but the position of the R¹ group seemed to impose some effect on the enantioselectivity due to the fact that C6-substituted substrates provided higher ee value than their C5- or C7-substituted analogues (entries 3–5 vs 1, 2 and 6, 7). As for the R² group, it could be successfully changed from an ester group to a benzoyl group or a cyano group, which gave the corresponding bis-spirooxindoles with perfected enantioselectivities (96%–98% ee, entries 8 and 9). These results demonstrated that this reaction had a wide substrate scope, which could be readily utilized for the enantioselective synthesis of such bis-spirooxindoles with structural diversity.

Finally, in order to examine the applicability of *N*-unprotected isatin in the reaction, we employed *N*-H isatin **1n** as a substrate to the reactions with amino ester **2** and methyleneindolinone **3j** (eq 3). Gratifyingly, this *N*-unpro-



TECTED isatin successfully participated in the desired reaction to afford the corresponding bis-spirooxindole **4nj** in a high yield of 96% and an excellent stereoselectivity of >95:5 dr and 90% ee, which indicated that the *N*-H group of isatins had no remarkable effect on the reactivity and the stereoselectivity. In addition, we also investigated the effect of nitro functionality as a strong electron-withdrawing group on the reaction by using *C*5-nitro-substituted isatin **1o** as a reactant (eq 4). Indeed, the



nitro group imposed an obviously negative effect on the stereoselectivity because the product **4oj** was generated in a low diastereo- and enantioselectivity albeit with a high yield.

The absolute configuration of bis-spirooxindole **4aa** was unambiguously determined to be (3*R*,4*S*,5*R*) by X-ray single-crystal analysis (>99% ee after recrystallization).¹² The relative and absolute configurations of other bis-spirooxindole products **4** were assigned by analogy. Based on the experimental results and previous report^{10b} on the activation mode of bis-PA to dipoles as well as dipolarophiles in catalytic asymmetric 1,3-DCs, we suggested a possible reaction pathway and activation mode. As illustrated in Scheme 1, bis-PA simultaneously generated two hydrogen bonds with the amide C=O group of methyleneindolinones and the N–H group of azomethine ylides, which were produced in situ by the condensation of isatins and amino ester in the presence of bis-PA. Thus, bis-PA acted as a bifunctional catalyst, and this dual-activation mode of the catalyst to the two reactants facilitated the subsequent enantioselective [3 + 2] cycloaddition. The reaction may largely undergo a concerted pathway because no intermediate Michael addition product was observed at all. Obviously, the regioselectivity of the 1,3-DC was controlled by the amide C=O group rather than the ester group of methyleneindolinones, which may also be ascribed to the hydrogen-bonding interaction between the catalyst and the amide C=O group. Bis-PA was supposed to form an intramolecular hydrogen-bond between the two acidic functionalities, thus making it easy for this catalyst to hold bulky isatin-derived substrates and impose the effect of enantioselective induction.^{10b} Therefore, the unique structure of bis-PA and its powerful dual hydrogen-bonding activation mode contributed greatly to the observed excellent stereoselectivity of the reaction.

CONCLUSIONS

In summary, we have achieved the first catalytic enantioselective construction of the 3,3'-pyrrolidinyl-dispirooxindole scaffold, which takes use of organocatalytic asymmetric 1,3-

DCs of isatin-derived azomethine ylides with methyleneindolinones. By using this strategy, a variety of structurally complex bis-spirooxindoles containing three adjacent and two quaternary stereogenic centers have been efficiently synthesized in generally high yields (up to 99%) and excellent diastereo- and enantioselectivities (up to >95:5 dr, 98% ee). The experimentally observed excellent stereoselectivity may largely be attributed to the unique structure of bis-PA catalyst and its powerful dual hydrogen-bonding activation mode to the reactants. This reaction not only provides a robust method for diastereo- and enantioselective construction of the chiral bis-spirooxindole scaffold but also greatly expands the application of catalytic asymmetric 1,3-DCs of azomethine ylides in synthesizing biologically important spirooxindoles with optical purity.

EXPERIMENTAL SECTION

General Information. ¹H and ¹³C NMR spectra were measured at 400 and 100 MHz, respectively. The solvent used for NMR spectroscopy was CDCl₃, using tetramethylsilane as the internal reference. HRMS (ESI) was determined using an HRMS/MS instrument. Enantiomeric excesses (ee) were determined by chiral high-performance liquid chromatography (chiral HPLC). The chiral columns used for the determination of enantiomeric excesses by chiral HPLC was Chiralpak AD-H and IA. Optical rotation values were measured with instruments operating at λ = 589 nm, corresponding to the sodium D line at the temperatures indicated. The X-ray source used for the single-crystal X-ray diffraction analysis of compound **4aa** was Cu Kα (λ = 1.54178), and the thermal ellipsoid was drawn at the 30% probability level. Analytical-grade solvents for column chromatography and commercially available reagents were used as received. All commercially available starting materials were used directly. Substrates **1** and **3** were synthesized according to the literature methods.^{10b,13} Catalyst **6** was prepared according to the procedures reported in the literature.^{7c}

Typical Procedure for the Catalytic Enantioselective Synthesis of 3,3'-Pyrrolidinyl-dispirooxindoles 4. After a solution of isatins **1** (0.12 mmol), amino ester **2** (0.1 mmol), the catalyst **6** (0.015 mmol), and 3 Å molecular sieves (100 mg) in chloroform (0.5 mL) was stirred at room temperature for 30 min, the solution of methyleneindolinones **3** (0.15 mol) in chloroform (0.5 mL) was added. After being stirred at 50 °C for 36 h, the reaction mixture was filtered to remove the molecular sieves, and the solid powder was washed with ethyl acetate. The resultant solution was concentrated under the reduced pressure to give the residue, which was purified through flash column chromatography on silica gel to afford bis-spirooxindole products **4**.

Compound 4aa: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield 99% (61.9 mg); >95:5 dr; white solid; mp 148–149 °C; [α]_D²⁰ = +63.5 (c 0.89, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.87 (d, *J* = 7.6 Hz, 1H), 7.44 (d, *J* = 7.5 Hz, 1H), 7.27 (d, *J* = 8.6 Hz, 1H), 7.21–7.15 (m, 3H), 7.13 (t, *J* = 7.7 Hz, 1H), 7.07 (t, *J* = 7.6 Hz, 1H), 7.00 (t, *J* = 7.6 Hz, 1H), 6.88–6.80 (m, 2H), 6.69 (d, *J* = 7.7 Hz, 1H), 6.54 (d, *J* = 7.8 Hz, 1H), 5.29 (s, 1H), 5.21 (s, 1H), 4.99 (d, *J* = 15.6 Hz, 1H), 4.55–4.39 (m, 2H), 4.38–4.29 (m, 2H), 4.14 (d, *J* = 15.6 Hz, 1H), 3.81–3.66 (m, 2H), 2.88 (s, 3H), 1.33 (t, *J* = 7.1 Hz, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.3, 173.1, 171.7, 169.2, 167.3, 142.7, 141.4, 137.8, 134.5, 129.8, 129.5, 129.2, 128.7, 127.9, 127.8, 127.4, 126.9, 125.5, 124.4, 122.1, 121.9, 108.7, 107.6, 74.9, 74.2, 64.8, 63.1, 63.0, 60.9, 58.9, 43.5, 26.7, 14.0, 13.8, 13.4; IR (KBr) 3450, 3270, 3003, 2977, 1731, 1647, 1557, 1541, 1473, 757 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₅N₃O₈ + Na)⁺ requires *m/z* 648.2316, found *m/z* 648.2301; ee 86%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 9.58 min (minor), *t*_R = 18.05 min (major).

Compound 4ba: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield 95% (64.7 mg);

>95:5 dr; white solid; mp 99–101 °C; $[\alpha]_{\text{D}}^{20} = +114.3$ (*c* 1.25, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, *J* = 7.2 Hz, 1H), 7.43 (d, *J* = 7.1 Hz, 1H), 7.30–7.26 (m, 1H), 7.19 (d, *J* = 8.3 Hz, 2H), 7.17–7.11 (m, 1H), 7.10–7.04 (m, 1H), 7.03–6.96 (m, 1H), 6.80 (d, *J* = 8.3 Hz, 2H), 6.69 (d, *J* = 7.7 Hz, 1H), 6.59 (d, *J* = 7.7 Hz, 1H), 5.22 (s, 1H), 4.94 (d, *J* = 15.4 Hz, 1H), 4.62 (s, 1H), 4.54–4.41 (m, 2H), 4.37–4.27 (m, 2H), 4.11 (d, *J* = 15.4 Hz, 1H), 3.79–3.65 (m, 2H), 2.87 (s, 3H), 1.36–1.30 (m, 6H), 1.27 (s, 9H), 0.69 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.2, 172.7, 171.7, 169.2, 167.3, 150.2, 142.7, 141.6, 132.7, 129.4, 129.0, 128.9, 127.9, 127.2, 127.0, 125.4, 124.6, 121.9, 121.8, 108.9, 107.3, 74.8, 74.2, 64.9, 63.1, 63.0, 60.9, 58.9, 43.5, 34.4, 31.3, 26.5, 14.0, 13.8, 13.4; IR (KBr) 3429, 3277, 3060, 2971, 1732, 1612, 1569, 1493, 758 cm⁻¹; ESI FTMS exact mass calcd for (C₃₉H₄₃N₃O₈ + Na)⁺ requires *m/z* 704.2947, found *m/z* 704.2947; enantiomeric excess 88%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 7.22 min (minor), *t*_R = 9.68 min (major).

Compound 4ca: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 89% (62.6 mg); >95:5 dr; white solid; mp 128–129 °C; $[\alpha]_{\text{D}}^{20} = +83.2$ (*c* 1.17, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 7.5 Hz, 1H), 7.45 (d, *J* = 7.3 Hz, 1H), 7.30–7.26 (m, 3H), 7.16–7.09 (m, 1H), 7.08–6.97 (m, 2H), 6.69 (t, *J* = 8.3 Hz, 3H), 6.48 (d, *J* = 7.7 Hz, 1H), 5.15 (s, 1H), 4.89 (d, *J* = 15.8 Hz, 1H), 4.59–4.38 (m, 3H), 4.38–4.28 (m, 2H), 4.11 (d, *J* = 15.8 Hz, 1H), 3.78–3.70 (m, 2H), 2.88 (s, 3H), 1.35–1.29 (m, 6H), 0.72 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.4, 173.2, 171.5, 169.1, 167.2, 142.8, 141.3, 134.7, 131.6, 129.5, 129.1, 128.9, 128.8, 127.7, 127.3, 124.4, 122.1, 122.0, 121.3, 108.6, 107.3, 75.0, 74.3, 64.7, 63.0, 60.9, 58.7, 43.2, 26.5, 14.0, 13.8, 13.4; IR (KBr) 3446, 3281, 3053, 2978, 1727, 1610, 1578, 1488, 765 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄BrN₃O₈ + Na)⁺ requires *m/z* 726.1421, found *m/z* 726.1419; enantiomeric excess: 84%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 10.71 min (minor), *t*_R = 14.20 min (major).

Compound 4da: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 85% (56.0 mg); >95:5 dr; white solid; mp 119–121 °C; $[\alpha]_{\text{D}}^{20} = +112.4$ (*c* 1.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 7.5 Hz, 1H), 7.45 (d, *J* = 7.4 Hz, 1H), 7.29–7.26 (m, 1H), 7.19–7.10 (m, 3H), 7.08–6.98 (m, 2H), 6.80 (d, *J* = 11.4 Hz, 2H), 6.73 (d, *J* = 7.7 Hz, 1H), 6.52 (d, *J* = 7.7 Hz, 1H), 5.21 (s, 1H), 4.98 (d, *J* = 15.8 Hz, 1H), 4.59 (s, 1H), 4.52–4.41 (m, 2H), 4.36–4.28 (m, 2H), 4.08 (d, *J* = 15.8 Hz, 1H), 3.78–3.68 (m, 2H), 2.93 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 6H), 0.69 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.3, 173.1, 171.7, 169.1, 167.2, 142.7, 141.3, 137.8, 134.4, 129.8, 129.5, 129.2, 128.7, 127.8, 127.3, 126.9, 125.4, 124.4, 122.1, 121.9, 108.7, 107.6, 74.9, 74.2, 64.8, 63.1, 63.0, 60.9, 58.8, 43.5, 26.7, 14.0, 13.8, 13.4; IR (KBr) 3445, 3273, 2979, 2925, 1716, 1608, 1576, 1489, 754 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄ClN₃O₈ + Na)⁺ requires *m/z* 682.1926, found *m/z* 682.1917; enantiomeric excess 88%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 9.98 min (minor), *t*_R = 17.02 min (major).

Compound 4ea: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 86% (56.7 mg); >95:5 dr; white solid; mp 97–98 °C; $[\alpha]_{\text{D}}^{20} = +81.0$ (*c* 0.92, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 7.5 Hz, 1H), 7.48 (d, *J* = 7.3 Hz, 1H), 7.32–7.27 (m, 1H), 7.25–7.20 (m, 1H), 7.17–7.10 (m, 2H), 7.06–6.95 (m, 3H), 6.67 (d, *J* = 7.7 Hz, 1H), 6.52 (d, *J* = 7.7 Hz, 1H), 6.46 (d, *J* = 7.6 Hz, 1H), 5.21 (s, 1H), 4.98 (d, *J* = 16.5 Hz, 1H), 4.60 (s, 1H), 4.53–4.28 (m, 5H), 3.81–3.69 (m, 2H), 2.97 (s, 3H), 1.37–1.29 (m, 6H), 0.71 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.4, 173.3, 171.6, 169.1, 167.2, 142.7, 141.4, 132.8, 132.7, 129.6, 129.5, 129.1, 128.9, 128.6, 127.8, 127.7, 127.3, 126.8, 124.4, 122.1, 122.0, 108.9, 107.3, 74.9, 74.3, 64.7, 63.1, 63.0, 60.9, 59.0, 41.6, 26.6, 14.0, 13.8, 13.4; IR (KBr) 3477, 3298, 2976, 2923, 1728, 1612, 1491, 755 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄ClN₃O₈ + Na)⁺ requires *m/z* 682.1926, found *m/z* 682.1926; enantiomeric excess 86%, determined by HPLC (Daicel Chiralpak IA, hexane/2-

propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 8.43 min (minor), *t*_R = 12.28 min (major).

Compound 4fa: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 88% (53.8 mg); >95:5 dr; white solid; mp 173–175 °C; $[\alpha]_{\text{D}}^{20} = -12.8$ (*c* 1.03, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.72–7.65 (m, 1H), 7.53–7.47 (m, 1H), 7.38–7.31 (m, 2H), 7.30–7.27 (m, 1H), 7.25–7.19 (m, 1H), 7.15–7.09 (m, 1H), 7.07–7.01 (m, 1H), 7.01–6.96 (m, 1H), 6.95–6.91 (m, 2H), 6.67 (d, *J* = 7.6 Hz, 1H), 6.53 (d, *J* = 7.5 Hz, 1H), 5.10 (s, 1H), 4.55 (s, 1H), 4.51–4.40 (m, 2H), 4.40–4.29 (m, 2H), 3.82–3.74 (m, 2H), 2.96 (s, 3H), 1.38–1.29 (m, 6H), 0.77 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.9, 172.8, 171.3, 169.0, 167.3, 143.0, 142.6, 133.9, 129.4, 129.4, 129.1, 128.6, 127.9, 127.1, 127.1, 126.4, 124.2, 122.3, 122.0, 109.0, 107.4, 75.6, 74.9, 64.9, 63.0, 62.9, 60.9, 58.2, 26.5, 14.0, 13.9, 13.5; IR (KBr) 3461, 3276, 3056, 2980, 1718, 1610, 1594, 1498, 751 cm⁻¹; ESI FTMS exact mass calcd for (C₃₄H₃₃N₃O₈ + Na)⁺ requires *m/z* 634.2160, found *m/z* 634.2159; enantiomeric excess 90%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 85/15, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 15.19 min (minor), *t*_R = 17.14 min (major).

Compound 4ga: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 83% (45.6 mg); >95:5 dr; white solid; mp 184–186 °C; $[\alpha]_{\text{D}}^{20} = +103.1$ (*c* 0.91, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 7.6 Hz, 1H), 7.45 (d, *J* = 7.5 Hz, 1H), 7.25–7.19 (m, 2H), 7.09–6.98 (m, 2H), 6.70–6.59 (m, 2H), 5.19 (s, 1H), 4.52 (s, 1H), 4.49–4.36 (m, 2H), 4.35–4.22 (m, 2H), 3.76–3.65 (m, 2H), 2.95 (s, 3H), 2.88 (s, 3H), 1.33–1.27 (m, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.4, 172.7, 171.6, 169.0, 167.2, 142.7, 142.5, 129.5, 129.0, 128.7, 127.3, 127.2, 124.2, 121.7, 121.6, 108.1, 107.4, 74.7, 73.9, 64.3, 63.0, 62.9, 60.9, 59.4, 26.6, 26.5, 13.9, 13.8, 13.4; IR (KBr) 3425, 3279, 2958, 2924, 2359, 1721, 1611, 1539, 1495, 766 cm⁻¹; ESI FTMS exact mass calcd for (C₂₉H₃₁N₃O₈ + Na)⁺ requires *m/z* 572.2003, found *m/z* 572.2031; enantiomeric excess 80%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 8.27 min (minor), *t*_R = 10.86 min (major).

Compound 4ha: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 93% (59.8 mg); >95:5 dr; white solid; mp 163–164 °C; $[\alpha]_{\text{D}}^{20} = +88.0$ (*c* 1.13, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.84 (d, *J* = 7.5 Hz, 1H), 7.28 (d, *J* = 7.7 Hz, 1H), 7.25–7.23 (m, 1H), 7.21–7.16 (m, 3H), 7.07 (t, *J* = 7.5 Hz, 1H), 6.86–6.78 (m, 3H), 6.71 (d, *J* = 7.7 Hz, 1H), 6.47–6.41 (m, 1H), 5.17 (s, 1H), 4.98 (d, *J* = 15.7 Hz, 1H), 4.62 (s, 1H), 4.53–4.40 (m, 2H), 4.39–4.28 (m, 2H), 4.11 (d, *J* = 15.7 Hz, 1H), 3.77–3.67 (m, 2H), 2.91 (s, 3H), 1.37–1.30 (m, 6H), 0.71 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.1, 172.7, 171.62, 169.1, 167.1, 158.5 (*J* = 239.2 Hz), 142.7, 137.5, 137.4, 135.3, 129.8, 129.7, 129.3, 128.9, 128.6, 127.6, 127.0, 124.3, 122.0, 115.8 (*J* = 8.3 Hz), 115.5 (*J* = 10.8 Hz), 109.3 (*J* = 8.0 Hz), 107.4, 74.8, 74.5, 64.8, 63.2, 63.1, 61.0, 58.8, 44.0, 26.6, 14.0, 13.9, 13.4; IR (KBr) 3429, 3278, 3084, 2924, 1718, 1636, 1559, 1507, 787 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄FN₃O₈ + Na)⁺ requires *m/z* 666.2222, found *m/z* 666.2258; enantiomeric excess 86%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t*_R = 8.28 min (minor), *t*_R = 13.84 min (major).

Compound 4ia: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 75% (49.1 mg); >95:5 dr; white solid; mp 150–151 °C; $[\alpha]_{\text{D}}^{20} = +69.9$ (*c* 0.72, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.84 (d, *J* = 7.1 Hz, 1H), 7.34 (d, *J* = 8.5 Hz, 1H), 7.25–7.23 (m, 1H), 7.22–7.12 (m, 3H), 7.10–7.00 (m, 1H), 6.85–6.79 (m, 2H), 6.68 (d, *J* = 7.7 Hz, 1H), 6.50–6.45 (m, 1H), 6.09 (d, *J* = 2.3 Hz, 1H), 5.16 (s, 1H), 5.06–4.86 (m, 1H), 4.55–4.39 (m, 3H), 4.37–4.27 (m, 2H), 4.11 (d, *J* = 15.7 Hz, 1H), 3.79–3.70 (m, 2H), 3.69 (s, 3H), 2.90 (s, 3H), 1.35–1.30 (m, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.6, 173.3, 171.7, 169.1, 167.4, 160.6, 143.0, 142.8, 135.6, 129.0, 128.8, 128.5, 128.1, 127.4, 127.0, 124.6, 121.9, 119.6, 107.3, 105.3, 96.9, 74.8, 74.0, 64.8, 62.7, 60.9, 58.7, 55.2, 43.8, 26.5, 14.0, 13.9, 13.4; IR (KBr) 3476, 3352, 3065, 2957, 1732, 1622, 1578, 1504, 797 cm⁻¹; ESI FTMS exact mass calcd for (C₃₆H₃₇N₃O₉ + Na)⁺ requires *m/z* 678.2422, found *m/z*

z 678.2458; enantiomeric excess 82%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 12.69 min (minor), *t_R* = 25.04 min (major).

Compound 4ja: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 90% (59.3 mg); >95:5 dr; white solid; mp 159–160 °C; $[\alpha]_{\text{D}}^{20} = +88.3$ (*c* 0.92, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, *J* = 7.1 Hz, 1H), 7.37 (d, *J* = 8.2 Hz, 1H), 7.30–7.26 (m, 1H), 7.22–7.16 (m, 3H), 7.09–7.02 (m, 1H), 7.00–6.94 (m, 1H), 6.84–6.78 (m, 2H), 6.70 (d, *J* = 7.7 Hz, 1H), 6.52 (d, *J* = 1.8 Hz, 1H), 5.16 (s, 1H), 4.96 (d, *J* = 15.7 Hz, 1H), 4.55 (d, *J* = 15.5 Hz, 1H), 4.52–4.39 (m, 2H), 4.37–4.26 (m, 2H), 4.11 (d, *J* = 15.7 Hz, 1H), 3.80–3.67 (m, 2H), 2.90 (s, 3H), 1.37–1.26 (m, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.2, 173.0, 171.6, 169.1, 167.2, 142.8, 142.7, 135.2, 135.0, 129.3, 128.8, 128.7, 128.3, 127.6, 127.0, 126.3, 124.3, 122.1, 122.0, 109.5, 107.5, 74.8, 74.0, 64.8, 63.1, 63.1, 61.0, 58.7, 43.9, 26.6, 14.0, 13.9, 13.4; IR (KBr) 3441, 3282, 2974, 2936, 2362, 1727, 1610, 1560, 1492, 753 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄ClN₃O₈ + Na)⁺ requires *m/z* 682.1926, found *m/z* 682.1912; enantiomeric excess: 90%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm): *t_R* = 9.56 min (minor), *t_R* = 17.25 min (major).

Compound 4ka: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 90% (63.3 mg); >95:5 dr; white solid; mp 148–149 °C; $[\alpha]_{\text{D}}^{20} = +102.6$ (*c* 0.94, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.85–7.77 (m, 1H), 7.31 (d, *J* = 8.2 Hz, 1H), 7.29–7.26 (m, 1H), 7.23–7.17 (m, 3H), 7.15–7.10 (m, 1H), 7.09–7.03 (m, 1H), 6.85–6.77 (m, 2H), 6.70 (d, *J* = 7.7 Hz, 1H), 6.67 (d, *J* = 1.7 Hz, 1H), 5.16 (s, 1H), 4.96 (d, *J* = 15.7 Hz, 1H), 4.51–4.40 (m, 2H), 4.37–4.25 (m, 2H), 4.11 (d, *J* = 15.7 Hz, 1H), 3.83–3.68 (m, 2H), 2.91 (s, 3H), 1.32 (t, *J* = 7.1 Hz, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.2, 172.8, 171.6, 169.0, 167.1, 142.9, 142.7, 135.0, 129.2, 128.8, 128.6, 128.6, 127.6, 126.9, 126.8, 124.9, 124.2, 123.2, 122.1, 112.2, 107.4, 74.8, 74.0, 64.7, 63.1, 63.1, 60.9, 58.7, 43.8, 26.6, 14.0, 13.8, 13.4; IR (KBr) 3417, 3339, 3021, 2938, 1728, 1607, 1587, 1489, 754 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄BrN₃O₈ + Na)⁺ requires *m/z* 726.1421, found *m/z* 726.1427; enantiomeric excess 88%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 9.56 min (minor), *t_R* = 18.01 min (major).

Compound 4la: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 82% (56.8 mg); >95:5 dr; white solid; mp 141–143 °C; $[\alpha]_{\text{D}}^{20} = +104.7$ (*c* 1.03, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.83–7.76 (m, 1H), 7.73 (d, *J* = 7.0 Hz, 1H), 7.53 (d, *J* = 7.5 Hz, 1H), 7.25–7.19 (m, 1H), 7.17–7.12 (m, 1H), 7.13–7.08 (m, 3H), 7.07–7.00 (m, 1H), 6.68 (d, *J* = 7.7 Hz, 1H), 6.66–6.61 (m, 2H), 5.26 (s, 1H), 5.03 (t, *J* = 21.7 Hz, 1H), 4.69 (t, *J* = 13.5 Hz, 2H), 4.55–4.40 (m, 2H), 4.39–4.25 (m, 2H), 3.79–3.64 (m, 2H), 2.96 (s, 3H), 1.37–1.29 (m, 6H), 0.70 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 174.7, 172.7, 171.8, 169.2, 166.9, 142.6, 139.8, 135.9, 131.3, 131.1, 129.2, 129.0, 128.0, 127.6 (*J* = 6.1 Hz), 126.6, 125.6, 124.6, 124.0, 122.1, 121.9, 121.3, 112.6 (*J* = 32.7 Hz), 107.5, 74.6, 72.6, 64.9, 63.2, 63.1, 61.0, 59.2, 45.9, 45.9, 26.6, 14.0, 13.8, 13.4; IR (KBr) 3487, 3279, 3061, 2983, 1732, 1612, 1597, 1498, 1096, 751 cm⁻¹; ESI FTMS exact mass calcd for (C₃₆H₃₄F₃N₃O₈ + Na)⁺ requires *m/z* 716.2190, found *m/z* 716.2191; enantiomeric excess: 88%, determined by HPLC (Daicel Chiralpak AD-H, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 6.15 min (minor), *t_R* = 10.10 min (major).

Compound 4ma: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 87% (55.9 mg); >95:5 dr; white solid; mp 115–116 °C; $[\alpha]_{\text{D}}^{20} = +110.2$ (*c* 0.95, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.91–7.84 (m, 1H), 7.30–7.26 (m, 1H), 7.26–7.23 (m, 1H), 7.20–7.14 (m, 3H), 7.10–7.05 (m, 1H), 6.99–6.91 (m, 2H), 6.90–6.86 (m, 2H), 6.67 (d, *J* = 7.6 Hz, 1H), 5.20 (s, 1H), 4.95 (d, *J* = 15.2 Hz, 1H), 4.56–4.39 (m, 3H), 4.37–4.25 (m, 2H), 3.76–3.63 (m, 2H), 2.84 (s, 3H), 1.36–1.29 (m, 6H), 0.67 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.6 (*J* = 38.6 Hz), 171.8, 169.1, 167.1, 148.6, 146.2, 142.6, 136.8, 131.0, 129.1 (*J* = 15.9 Hz), 128.3, 128.2, 127.5, 127.3, 124.2, 123.4 (*J* = 3.2

Hz), 122.4 (*J* = 6.3 Hz), 121.9, 117.6 (*J* = 19.2 Hz), 107.4, 74.7, 74.3, 74.2, 65.0, 63.2, 63.1, 60.9, 58.9, 45.3, 45.2, 26.5, 13.9, 13.8, 13.4; IR (KBr) 3457, 3387, 3079, 2976, 1718, 1629, 1608, 1489, 771 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₄FN₃O₈ + Na)⁺ requires *m/z* 666.2222, found *m/z* 666.2217; enantiomeric excess 92%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm): *t_R* = 7.82 min (minor), *t_R* = 15.82 min (major).

Compound 4ab: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 90% (57.5 mg); >95:5 dr; white solid; mp 81–82 °C; $[\alpha]_{\text{D}}^{20} = +88.4$ (*c* 0.95, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.61 (s, 1H), 7.50–7.46 (m, 1H), 7.21–7.16 (m, 3H), 7.14–7.09 (m, 1H), 7.08–7.04 (m, 1H), 7.03–6.97 (m, 1H), 6.86–6.79 (m, 2H), 6.59 (d, *J* = 7.9 Hz, 1H), 6.50 (d, *J* = 7.6 Hz, 1H), 5.13 (s, 1H), 5.06 (d, *J* = 15.8 Hz, 1H), 4.59–4.42 (m, 3H), 4.41–4.30 (m, 2H), 4.15 (d, *J* = 15.8 Hz, 1H), 3.87–3.75 (m, 2H), 2.90 (s, 3H), 2.32 (s, 3H), 1.39–1.32 (m, 6H), 0.77 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.6, 173.4, 171.5, 169.0, 167.4, 141.9, 140.6, 135.6, 131.4, 129.5, 129.4, 129.3, 128.5, 127.6, 127.3, 127.0, 126.8, 124.2, 121.9, 108.8, 107.0, 75.3, 74.4, 64.6, 62.3, 62.9, 60.8, 58.4, 43.7, 26.4, 21.3, 14.0, 13.9, 13.4; IR (KBr) 3432, 3278, 2981, 2937, 1730, 1613, 1500, 1470, 1357, 750 cm⁻¹; ESI FTMS exact mass calcd for (C₃₆H₃₇N₃O₈ + Na)⁺ requires *m/z* 662.2473, found *m/z* 662.2471; enantiomeric excess 88%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 80/20, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm): *t_R* = 10.97 min (minor), *t_R* = 13.22 min (major).

Compound 4mc: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 64% (43.3 mg); >95:5 dr; white solid; mp 163–165 °C; $[\alpha]_{\text{D}}^{20} = +59.5$ (*c* 0.87, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, *J* = 1.9 Hz, 1H), 7.31–7.27 (m, 1H), 7.25 (d, *J* = 2.0 Hz, 1H), 7.21–7.15 (m, 3H), 7.01–6.86 (m, 4H), 6.58 (d, *J* = 8.3 Hz, 1H), 5.14 (s, 1H), 4.96 (d, *J* = 15.2 Hz, 1H), 4.57–4.26 (m, 6H), 3.88–3.71 (m, 2H), 2.81 (s, 3H), 1.34 (q, *J* = 7.2 Hz, 6H), 0.77 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.4 (*J* = 5.1 Hz), 171.4, 169.0, 166.8, 147.4 (*J* = 242.2 Hz), 141.2, 136.7, 130.7, 129.4, 128.9, 128.3, 128.2 (*J* = 8.7 Hz), 127.5, 127.4, 127.4, 127.3, 125.8, 123.3 (*J* = 3.2 Hz), 122.5 (*J* = 6.3 Hz), 117.7 (*J* = 9.6 Hz), 108.2, 74.5, 74.1, 74.0, 64.5, 63.3, 63.2, 61.1, 58.5, 45.3, 45.2, 26.6, 13.9, 13.8, 13.5; IR (KBr) 3442, 3288, 3061, 2935, 1731, 1612, 1567, 1489, 1473, 737 cm⁻¹; ESI FTMS exact mass calcd for (C₃₅H₃₃ClFN₃O₈ + Na)⁺ requires *m/z* 700.1832, found *m/z* 700.1877; enantiomeric excess 86%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 7.57 min (minor), *t_R* = 9.39 min (major).

Compound 4md: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 90% (60.6 mg); >95:5 dr; white solid; mp 151–153 °C; $[\alpha]_{\text{D}}^{20} = +128.9$ (*c* 1.07, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.77 (d, *J* = 8.5 Hz, 1H), 7.25–7.21 (m, 1H), 7.21–7.14 (m, 3H), 7.04–6.85 (m, 4H), 6.61–6.51 (m, 1H), 6.24 (d, *J* = 2.3 Hz, 1H), 5.13 (s, 1H), 4.98 (d, *J* = 15.2 Hz, 1H), 4.60 (s, 1H), 4.54–4.38 (m, 3H), 4.37–4.25 (m, 2H), 3.80 (s, 3H), 3.80–3.59 (m, 2H), 2.81 (s, 3H), 1.40–1.25 (m, 6H), 0.74 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.0 (*J* = 6.7 Hz), 171.7, 169.2, 167.2, 160.7, 147.3 (*J* = 241.9 Hz), 144.0, 136.8, 131.1 (*J* = 2.1 Hz), 129.9, 128.3, 128.2, 127.4, 127.4, 127.3, 123.2 (*J* = 3.1 Hz), 122.4 (*J* = 6.3 Hz), 117.5 (*J* = 19.3 Hz), 115.9, 104.9, 95.8, 74.6, 74.3, 74.2, 64.7, 63.1, 63.0, 60.9, 58.8, 55.3, 45.3, 45.2, 26.5, 13.9, 13.8, 13.5; IR (KBr) 3419, 3292, 2961, 2851, 1732, 1651, 1597, 1471, 1280, 702 cm⁻¹; ESI FTMS exact mass calcd for (C₃₆H₃₆FN₃O₉ + Na)⁺ requires *m/z* 696.2328, found *m/z* 696.2350; enantiomeric excess: 92%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 10.09 min (minor), *t_R* = 18.46 min (major).

Compound 4me: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 87% (62.7 mg); >95:5 dr; white solid; mp 138–140 °C; $[\alpha]_{\text{D}}^{20} = +135.5$ (*c* 1.26, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 8.2 Hz, 1H), 7.25–7.19 (m, 4H), 7.19–7.15 (m, 1H), 7.00–6.89 (m, 4H), 6.74 (d, *J* = 1.7 Hz, 1H), 5.14 (s, 1H), 4.93 (d, *J* = 15.1 Hz, 1H), 4.57–4.51 (m, 2H), 4.50–4.38 (m, 2H), 4.37–4.26 (m, 2H), 3.80–3.69 (m, 2H),

2.75 (s, 3H), 1.31 (t, $J = 7.1$ Hz, 6H), 0.75 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.4 ($J = 15.4$ Hz), 171.5, 169.2, 166.9, 148.4 ($J = 242.1$ Hz), 143.8, 136.7, 130.8 ($J = 2.0$ Hz), 130.4, 128.4, 128.1 ($J = 8.5$ Hz), 127.7, 127.6, 127.5, 124.7, 123.3 ($J = 3.2$ Hz), 123.1, 122.9, 122.5 ($J = 6.3$ Hz), 117.7 ($J = 19.3$ Hz), 111.1, 74.5, 74.2, 74.1, 64.7, 63.3, 63.2, 61.1, 58.6, 45.4, 45.3, 26.6, 14.0, 13.8, 13.5; IR (KBr) 3417, 3279, 3067, 2974, 1733, 1606, 1541, 1490, 737 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{35}\text{H}_{33}\text{BrFN}_3\text{O}_8 + \text{Na})^+$ requires m/z 744.1327, found m/z 744.1314; enantiomeric excess 94%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 6.66$ min (minor), $t_{\text{R}} = 12.55$ min (major).

Compound 4mf: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 85% (57.6 mg); >95:5 dr; white solid; mp 144–145 °C; $[\alpha]_{\text{D}}^{20} = +133.7$ (c 0.99, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.78 (d, $J = 8.2$ Hz, 1H), 7.25–7.17 (m, 4H), 7.05–7.00 (m, 1H), 6.99–6.88 (m, 4H), 6.60 (d, $J = 1.9$ Hz, 1H), 5.15 (s, 1H), 4.94 (d, $J = 15.1$ Hz, 1H), 4.54 (d, $J = 14.8$ Hz, 2H), 4.51–4.38 (m, 2H), 4.37–4.25 (m, 2H), 3.81–3.68 (m, 2H), 2.77 (s, 3H), 1.37–1.27 (m, 6H), 0.75 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.5 ($J = 29.4$ Hz), 171.5, 169.2, 166.9, 147.4 ($J = 242.1$ Hz), 143.7, 136.7, 135.0, 130.8 ($J = 2.0$ Hz), 130.1, 128.4, 128.1 ($J = 8.5$ Hz), 127.7, 127.6, 127.5, 123.3 ($J = 3.2$ Hz), 122.6 ($J = 6.2$ Hz), 122.5, 121.8, 117.7 ($J = 19.4$ Hz), 108.3, 74.5, 74.3, 74.2, 64.6, 63.3, 63.1, 61.1, 58.6, 45.4, 45.3, 26.6, 13.9, 13.8, 13.5; IR (KBr) 3437, 3279, 2975, 2926, 1732, 1610, 1557, 1495, 737 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{35}\text{H}_{33}\text{ClFN}_3\text{O}_8 + \text{Na})^+$ requires m/z 700.1832, found m/z 700.1826; enantiomeric excess 92%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 6.58$ min (minor), $t_{\text{R}} = 12.47$ min (major).

Compound 4ag: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 82% (52.4 mg); >95:5 dr; white solid; mp 143–145 °C; $[\alpha]_{\text{D}}^{20} = +118.7$ (c 1.02, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.76–7.66 (m, 1H), 7.43 (d, $J = 7.1$ Hz, 1H), 7.24–7.16 (m, 3H), 7.15–7.07 (m, 1H), 7.02–6.92 (m, 3H), 6.92–6.86 (m, 2H), 6.52 (d, $J = 7.7$ Hz, 1H), 5.23 (s, 1H), 5.01 (d, $J = 15.8$ Hz, 1H), 4.55–4.40 (m, 2H), 4.38–4.26 (m, 2H), 4.14 (d, $J = 15.8$ Hz, 1H), 3.84–3.68 (m, 2H), 3.19 (s, 3H), 2.44 (s, 3H), 1.36–1.29 (m, 6H), 0.72 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 174.3, 172.9, 171.8, 169.1, 167.4, 141.6, 140.5, 135.7, 132.9, 129.4, 128.5, 127.9, 127.4, 127.2, 126.9, 126.6, 125.2, 121.8, 121.7, 118.5, 108.9, 75.0, 74.5, 64.5, 63.1, 63.0, 60.8, 59.1, 43.8, 30.2, 19.3, 14.00, 13.9, 13.4; IR (KBr) 3477, 3289, 3029, 2973, 1732, 1609, 1488, 1468, 746 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{36}\text{H}_{37}\text{N}_3\text{O}_8 + \text{Na})^+$ requires m/z 662.2473, found m/z 662.2482; enantiomeric excess 88%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 10.54$ min (minor), $t_{\text{R}} = 18.50$ min (major).

Compound 4mh: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 62% (41.0 mg); >95:5 dr; pale yellow solid; mp 134–136 °C; $[\alpha]_{\text{D}}^{20} = +79.3$ (c 0.72, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.75–7.67 (m, 1H), 7.24–7.20 (m, 4H), 7.04–6.98 (m, 2H), 6.98–6.91 (m, 4H), 5.26 (d, $J = 30.1$ Hz, 1H), 5.00 (t, $J = 17.5$ Hz, 1H), 4.65–4.39 (m, 4H), 4.36–4.26 (m, 2H), 3.84–3.64 (m, 2H), 3.26–2.87 (m, 3H), 1.35–1.29 (m, 6H), 0.73 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.2 ($J = 29.0$ Hz), 171.7, 168.0 ($J = 215.9$ Hz), 148.5, 146.2 ($J = 10.6$ Hz), 136.7, 130.7, 130.7, 129.4 ($J = 8.3$ Hz), 128.5, 128.1 ($J = 8.6$ Hz), 127.6, 127.4 ($J = 1.5$ Hz), 127.0 ($J = 2.6$ Hz), 125.0 ($J = 3.2$ Hz), 123.4 ($J = 3.2$ Hz), 122.5 ($J = 6.3$ Hz), 122.2 ($J = 6.2$ Hz), 117.7 ($J = 19.3$ Hz), 117.0 ($J = 18.7$ Hz), 74.5, 74.3, 74.2, 65.2 ($J = 2.1$ Hz), 63.3, 63.1, 61.0, 59.0, 45.3, 45.3, 29.1, 29.0, 14.0, 13.8, 13.4; IR (KBr) 3487, 3281, 3068, 2982, 2361, 1732, 1629, 1598, 1488, 734 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{35}\text{H}_{33}\text{F}_2\text{N}_3\text{O}_8 + \text{Na})^+$ requires m/z 684.2128, found m/z 684.2150; enantiomeric excess 86%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 7.37$ min (minor), $t_{\text{R}} = 15.00$ min (major).

Compound 4mi: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 80% (54.1 mg); >95:5 dr; yellow solid; mp 155–157 °C; $[\alpha]_{\text{D}}^{20} = +168.4$ (c 0.91, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.88 (d, $J = 7.1$ Hz, 1H), 7.49–7.43 (m, 2H), 7.42–7.34 (m, 2H), 7.25–7.17 (m, 3H), 7.16–7.09 (m, 4H), 7.07–7.00 (m, 1H), 6.99–6.92 (m, 1H), 6.89–6.80 (m, 2H), 6.40 (d, $J = 7.6$ Hz, 1H), 6.31 (s, 1H), 4.93 (d, $J = 12.8$ Hz, 2H), 4.57–4.41 (m, 3H), 4.40–4.23 (m, 2H), 2.37 (s, 3H), 1.32–1.24 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 194.8, 172.4, 172.1 ($J = 10.8$ Hz), 169.5, 147.5 ($J = 242.0$ Hz), 141.7, 136.9, 136.8, 133.0, 131.7 ($J = 2.1$ Hz), 130.6, 129.1, 128.6, 128.3, 128.2, 128.1, 127.5, 127.4, 127.3, 123.4 ($J = 3.1$ Hz), 123.3, 122.4 ($J = 6.3$ Hz), 121.9, 117.4 ($J = 19.3$ Hz), 107.3, 75.4, 75.1, 75.0, 66.4, 63.2, 63.1, 61.2, 45.3, 45.2, 26.0, 13.9, 13.8; IR (KBr) 3453, 3276, 3062, 2927, 1732, 1611, 1597, 1492, 752 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{39}\text{H}_{34}\text{FN}_3\text{O}_7 + \text{Na})^+$ requires m/z 698.2273, found m/z 698.2272; enantiomeric excess 98%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 6.07$ min (minor), $t_{\text{R}} = 12.42$ min (major).

Compound 4mj: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 92% (54.8 mg); >95:5 dr; white solid; mp 93–95 °C; $[\alpha]_{\text{D}}^{20} = -224.8$ (c 0.84, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.61 (d, $J = 7.6$ Hz, 1H), 7.25–7.18 (m, 4H), 7.17–7.08 (m, 3H), 6.91–6.78 (m, 3H), 6.63 (d, $J = 7.8$ Hz, 1H), 4.96–4.79 (m, 2H), 4.71 (s, 1H), 4.50–4.37 (m, 4H), 4.10 (s, 1H), 3.05 (s, 3H), 1.42–1.33 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 174.7 ($J = 5.5$ Hz), 167.9, 166.7, 148.1, 145.7, 143.7, 136.1, 130.6 ($J = 4.4$ Hz), 130.1, 128.5, 127.5, 127.4, 125.8 ($J = 3.1$ Hz), 123.5, 123.4 ($J = 6.5$ Hz), 120.6 ($J = 1.6$ Hz), 120.1, 119.0 ($J = 19.4$ Hz), 116.1, 108.2, 75.0, 73.9, 73.8, 63.6, 63.2, 59.8, 45.3, 45.2, 42.3, 26.2, 14.1, 14.0; IR (KBr) 3467, 3255, 3021, 2928, 2246, 1732, 1612, 1531, 1472, 757 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{33}\text{H}_{29}\text{FN}_4\text{O}_6 + \text{Na})^+$ requires m/z 619.1963, found m/z 619.1954; enantiomeric excess 96%, determined by HPLC (Daicel Chiralpak IA, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 9.14$ min (minor), $t_{\text{R}} = 7.48$ min (major).

Compound 4nj: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 96% (48.6 mg); >95:5 dr; white solid; mp 222–223 °C; $[\alpha]_{\text{D}}^{20} = -129.1$ (c 0.91, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 8.25 (s, 1H), 7.81 (d, $J = 7.6$ Hz, 1H), 7.30–7.26 (m, 1H), 7.11–7.04 (m, 2H), 6.89–6.78 (m, 2H), 6.64 (d, $J = 7.8$ Hz, 1H), 4.70 (s, 1H), 4.50–4.33 (m, 4H), 4.07 (s, 1H), 3.05 (s, 3H), 1.36 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.9, 174.6, 167.8, 166.7, 146.5 ($J = 243.5$ Hz), 143.8, 130.3, 129.2 ($J = 12.8$ Hz), 127.3, 125.9 ($J = 3.3$ Hz), 123.6, 123.2 ($J = 5.8$ Hz), 120.5 ($J = 3.4$ Hz), 120.1, 117.8 ($J = 16.8$ Hz), 115.9, 108.2, 74.4, 74.3, 63.6, 63.5, 59.7, 42.0, 26.1, 14.0, 13.9; IR (KBr) 3450, 3310, 3066, 2914, 1731, 1647, 1557, 1530, 1489, 757 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{26}\text{H}_{23}\text{FN}_4\text{O}_6 + \text{Na})^+$ requires m/z 529.1499, found m/z 529.1487; enantiomeric excess: 90%, determined by HPLC (Daicel Chiralpak OD-H, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, $T = 30$ °C, 254 nm) $t_{\text{R}} = 8.46$ min (minor), $t_{\text{R}} = 13.23$ min (major).

Compound 4oj: flash column chromatography eluent, petroleum ether/ethyl acetate = 5/1; reaction time = 36 h; yield: 86% (54 mg); 55:45 dr (inseparable diastereomers); white solid; mp 115–116 °C; ^1H NMR (400 MHz, CDCl_3) δ 8.24 (d, $J = 2.2$ Hz, 1.2H), 8.22–8.17 (m, 1H), 8.04–8.00 (m, 1.2H), 7.96–7.92 (m, 1H), 7.68 (d, $J = 7.6$ Hz, 1.2H), 7.47 (d, $J = 2.3$ Hz, 1H), 7.40–7.27 (m, 8H), 7.25–7.18 (m, 4H), 6.99–6.87 (m, 4H), 6.73–6.64 (m, 2.4H), 6.58–6.43 (m, 2.4H), 5.49 (s, 1H), 5.16 (d, $J = 15.6$ Hz, 1H), 5.00 (d, $J = 15.9$ Hz, 1.2H), 4.81 (s, 1.2H), 4.60 (d, $J = 15.9$ Hz, 1.2H), 4.51–4.41 (m, 9.8H), 4.12 (s, 1.2H), 3.85 (s, 1H), 3.08 (s, 3.6H), 3.02 (s, 3H), 1.43–1.35 (m, 13.2H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.9, 175.2, 174.1, 170.2, 168.6, 167.5, 167.2, 166.6, 149.3, 149.1, 143.8, 143.5, 143.1, 142.7, 134.1, 133.6, 130.9, 130.4, 129.0, 128.9, 128.2, 128.0, 127.7, 127.5, 127.3, 127.2, 127.1, 124.2, 124.2, 123.6, 123.5, 123.2, 122.0, 121.0, 119.8, 115.9, 114.9, 109.2, 109.0, 108.8, 108.6, 73.3, 72.6, 71.7, 63.9, 63.8, 63.7, 63.5, 62.7, 59.5, 44.6, 44.1, 41.9, 41.4, 26.7, 26.3, 14.1, 14.0, 13.9, 13.9; IR (KBr) 3415, 3336, 3088, 2982, 1739, 1614, 1522, 1482, 1472, 758 cm^{-1} ; ESI FTMS exact mass calcd for $(\text{C}_{33}\text{H}_{29}\text{N}_5\text{O}_8 +$

Na)⁺ requires *m/z* 646.1913, found *m/z* 646.1919; enantiomeric excess of major diastereomer 32%, determined by HPLC (Daicel Chiralpak OD-H, hexane/2-propanol = 70/30, flow rate 1.0 mL/min, *T* = 30 °C, 254 nm) *t_R* = 27.28 min (minor), *t_R* = 34.46 min (major).

■ ASSOCIATED CONTENT

📄 Supporting Information

Details on screening of catalysts and solvents, characterization data (including ¹H and ¹³C NMR and HPLC spectra) for all products, and single-crystal data of product **4aa**. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00708.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: fshi@jnsu.edu.cn.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We are grateful for financial support from NSFC (21372002 and 21232007), Open Foundation of Jiangsu Key Laboratory (K201314), PAPD of Jiangsu Province, and the Qing Lan Project.

■ REFERENCES

(1) For some reviews, see: (a) Galliford, C. V.; Scheidt, K. A. *Angew. Chem., Int. Ed.* **2007**, *46*, 8748. (b) Singh, G. S.; Desta, Z. Y. *Chem. Rev.* **2012**, *112*, 6104. For some examples of natural products, see: (c) Jossang, A.; Jossang, P.; Hadi, H. A.; Sevenet, T.; Bodo, B. *J. Org. Chem.* **1991**, *56*, 6527. (d) Cui, C. B.; Kakeya, H.; Osada, H. *Tetrahedron* **1996**, *52*, 12651. For some examples of bioactive molecules, see: (e) Tripathy, R.; Reiboldt, A.; Messina, P. A.; Iqbal, M.; Singh, J.; Bacon, E. R.; Angeles, T. S.; Yang, S. X.; Albom, M. S.; Robinson, C.; Chang, H.; Ruggeri, B. A.; Mallamo, J. P. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 2158. (f) Nutan, G.; Modi, M.; Gupta, S. K.; Singh, R. K. *Eur. J. Med. Chem.* **2011**, *46*, 1181. (g) Yeung, B. K. S.; Zou, B.; Rottmann, M.; Lakshminarayana, S. B.; Ang, S. H.; Leong, S. Y.; Tan, J.; Wong, J.; Keller-Maerki, S.; Fischli, C.; Goh, A.; Schmitt, E. K.; Krastel, P.; Francotte, E.; Kuhlen, K.; Plouffe, D.; Henson, K.; Wagner, T.; Winzeler, E. A.; Petersen, F.; Brun, R.; Dartois, V.; Diagana, T. T.; Keller, T. H. *J. Med. Chem.* **2010**, *53*, 5155. (h) Murugan, R.; Anbazhagan, S.; Narayanan, S. S. *Eur. J. Med. Chem.* **2009**, *44*, 3272. (i) Rambabu, D.; Raja, G.; Sreenivas, B. Y.; Seerapu, G. P. K.; Kumar, K. L.; Deora, G. S.; Haldar, D.; Rao, M. V. B.; Pal, M. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1351.

(2) (a) Arun, Y.; Bhaskar, G.; Balachandran, C.; Ignacimuthu, S.; Perumal, P. T. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1839. (b) Velikorodov, A. V.; Ionova, V. A.; Degtyarev, O. V.; Sukhenko, L. T. *Pharm. Chem. J.* **2013**, *46*, 715. (c) Babu, A. R. S.; Raghunathan, R.; Mathivanan, N.; Omprabha, G.; Velmurugan, D.; Raghu, R. *Curr. Chem. Biol.* **2008**, *2*, 312.

(3) (a) Yeung, B. K. S.; Zou, B.; Rottmann, M.; Lakshminarayana, S. B.; Ang, S. H.; Leong, S. Y.; Tan, J.; Wong, J.; Keller-Maerki, S.; Fischli, C.; Goh, A.; Schmitt, E. K.; Krastel, P.; Francotte, E.; Kuhlen, K.; Plouffe, D.; Henson, K.; Wagner, T.; Winzeler, E. A.; Petersen, F.; Brun, R.; Dartois, V.; Diagana, T. T.; Keller, T. H. *J. Med. Chem.* **2010**, *53*, 5155. (b) Chinigo, G. M.; Paige, M.; Grindrod, S.; Hamel, E.; Dakshanamurthy, S.; Chruszcz, M.; Minor, W.; Brown, M. L. *J. Med. Chem.* **2008**, *51*, 4620.

(4) (a) Suman, K.; Srinu, L.; Thennarasu, S. *Org. Lett.* **2014**, *16*, 3732. (b) Lanka, S.; Thennarasu, S.; Perumal, P. T. *Tetrahedron Lett.* **2014**, *55*, 2585. (c) Dandia, A.; Jain, A. K.; Laxkar, A. K.; Bhati, D. S. *Tetrahedron* **2013**, *69*, 2062. (d) Liu, J.; Sun, H.; Liu, X.; Ouyang, L.; Kang, T.; Xie, Y.; Wang, X. *Tetrahedron Lett.* **2012**, *53*, 2336. (e) Jain, R.; Sharma, K.; Kumar, D. *Tetrahedron Lett.* **2012**, *53*, 1993. (f) Liu,

H.; Dou, G.; Shi, D. *J. Comb. Chem.* **2010**, *12*, 292. (g) Lakshmi, N. V.; Thirumurugan, P.; Perumal, P. T. *Tetrahedron Lett.* **2010**, *51*, 1064. (h) Shanmugam, P.; Viswambharan, B.; Selvakumar, K.; Madhavan, S. *Tetrahedron Lett.* **2008**, *49*, 2611. (i) Suresh Babu, A. R.; Raghunathan, R. *Tetrahedron* **2007**, *63*, 8010. (j) Jayashankaran, J.; Durga, R.; Manian, R.; Raghunathan, R. *Synth. Commun.* **2006**, *36*, 979–986.

(5) For diastereoselective synthesis of 3,3'-pyrrolidinyldispirooxindoles from chiral amino acids, see: (a) Xiao, J.-A.; Zhang, H.-G.; Liang, S.; Ren, J.-W.; Yang, H.; Chen, X.-Q. *J. Org. Chem.* **2013**, *78*, 11577. (b) Xu, Q.; Wang, D.; Wei, Y.; Shi, M. *ChemistryOpen* **2014**, *3*, 93. (c) Almansour, A. I.; Arumugam, N.; Kumar, R. S.; Periyasami, G.; Ghabbour, H. A.; Fun, H.-K. *Molecules* **2015**, *20*, 780.

(6) For early reports, see: (a) Longmire, J. M.; Wang, B.; Zhang, X. J. *Am. Chem. Soc.* **2002**, *124*, 13400. (b) Gothelf, A. S.; Gothelf, K. V.; Hazell, R. G.; Jorgensen, K. A. *Angew. Chem., Int. Ed.* **2002**, *41*, 4236. For some recent examples, see: (c) Teng, H.-L.; Yao, L.; Wang, C.-J. *J. Am. Chem. Soc.* **2014**, *136*, 4075. (d) Li, Q.-H.; Wei, L.; Wang, C.-J. *J. Am. Chem. Soc.* **2014**, *136*, 8685. (e) He, Z.-L.; Teng, H.-L.; Wang, C.-J. *Angew. Chem., Int. Ed.* **2013**, *52*, 2934. (f) Tong, M.-C.; Chen, X.; Tao, H.-Y.; Wang, C.-J. *Angew. Chem., Int. Ed.* **2013**, *52*, 12377. (g) Guo, H.; Liu, H.; Zhu, F.-L.; Na, R.; Jiang, H.; Wu, Y.; Zhang, L.; Li, Z.; Yu, H.; Wang, B.; Xiao, Y.; Hu, X.-P.; Wang, M. *Angew. Chem., Int. Ed.* **2013**, *52*, 12641. (h) Potowski, M.; Bauer, J. O.; Strohmann, C.; Antonchick, A. P.; Waldmann, H. *Angew. Chem., Int. Ed.* **2012**, *51*, 9512.

(7) For early reports, see: (a) Vicario, J. L.; Reboredo, S.; Badía, D.; Carrillo, L. *Angew. Chem., Int. Ed.* **2007**, *46*, 5168. (b) Ibrahim, I.; Rios, R.; Vesely, J.; Cordova, A. *Tetrahedron Lett.* **2007**, *48*, 6252. (c) Chen, X.-H.; Zhang, W.-Q.; Gong, L.-Z. *J. Am. Chem. Soc.* **2008**, *130*, 5652. (d) Liu, Y.-K.; Liu, H.; Du, W.; Yue, L.; Chen, Y.-C. *Chem.—Eur. J.* **2008**, *14*, 9873. (e) Xue, M.-X.; Zhang, X.-M.; Gong, L.-Z. *Synlett* **2008**, 691. For some recent examples: (f) Guo, C.; Song, J.; Gong, L.-Z. *Org. Lett.* **2013**, *15*, 2676. (g) Shi, F.; Xing, G.-J.; Tan, W.; Zhu, R.-Y.; Tu, S. *Org. Biomol. Chem.* **2013**, *11*, 1482.

(8) (a) Padwa, A.; Pearson, W. H. *Synthetic Applications of 1,3-Dipolar Cycloaddition Chemistry Toward Heterocycles and Natural Products*; John Wiley & Sons: New York, 2002. For some reviews, see: (b) Coldham, I.; Hufton, R. *Chem. Rev.* **2005**, *105*, 2765. (c) Nájera, J.; Sansano, M. *Angew. Chem., Int. Ed.* **2005**, *44*, 6272. (d) Pandey, G.; Banerjee, P.; Gadre, S. R. *Chem. Rev.* **2006**, *106*, 4484. (e) Nair, V.; Suja, T. D. *Tetrahedron* **2007**, *63*, 12247. (f) Pellissier, H. *Tetrahedron* **2007**, *63*, 3235. (g) Stanley, L. M.; Sibi, M. P. *Chem. Rev.* **2008**, *108*, 2887. (h) Marie, K.; Anita, R. M. *Chem. Soc. Rev.* **2010**, *39*, 845. (i) Adrio, J.; Carretero, J. C. *Chem. Commun.* **2011**, *47*, 6784. (j) Narayan, R.; Potowski, M.; Jia, Z.-J.; Antonchick, A. P.; Waldmann, H. *Acc. Chem. Res.* **2014**, *47*, 1296.

(9) For some reviews, see: (a) Akiyama, T. *Chem. Rev.* **2007**, *107*, 5744. (b) Terada, M. *Chem. Commun.* **2008**, 4097. (c) Terada, M. *Synthesis* **2010**, 1929. (d) Yu, J.; Shi, F.; Gong, L.-Z. *Acc. Chem. Res.* **2011**, *44*, 1156. (e) Parmar, D.; Sugiono, E.; Raja, S.; Rueping, M. *Chem. Rev.* **2014**, *114*, 9047.

(10) (a) Chen, X.-H.; Wei, Q.; Luo, S.-W.; Xiao, H.; Gong, L.-Z. *J. Am. Chem. Soc.* **2009**, *131*, 13819. (b) Shi, F.; Tao, Z.-L.; Luo, S.-W.; Tu, S.-J.; Gong, L.-Z. *Chem.—Eur. J.* **2012**, *18*, 6885. (c) Shi, F.; Zhu, R.-Y.; Liang, X.; Tu, S.-J. *Adv. Synth. Catal.* **2013**, *355*, 2447. (d) Wang, C.-S.; Zhu, R.-Y.; Zheng, J.; Shi, F.; Tu, S.-J. *J. Org. Chem.* **2015**, *80*, 512.

(11) (a) Tan, W.; Li, X.; Gong, Y.-X.; Ge, M.-D.; Shi, F. *Chem. Commun.* **2014**, *50*, 15901. (b) Zhang, Y.-C.; Zhao, J.-J.; Jiang, F.; Sun, S.-B.; Shi, F. *Angew. Chem., Int. Ed.* **2014**, *53*, 13912. (c) Zhao, J.-J.; Sun, S.-B.; He, S.-H.; Wu, Q.; Shi, F. *Angew. Chem., Int. Ed.* **2015**, *54*, 5460.

(12) CCDC 1056015 for **4aa**. See the Supporting Information for details.

(13) (a) Zheng, C.-W.; Lu, Y.-P.; Zhang, J.-K.; Chen, X.-K.; Chai, Z.; Ma, W.-Y.; Zhao, G. *Chem.—Eur. J.* **2010**, *16*, 5853. (b) Wei, Q.; Gong, L.-Z. *Org. Lett.* **2010**, *12*, 1008.